

Sialidase Inhibitors Related to Zanamivir. Further SAR Studies of 4-Amino-4*H*-pyran-2-carboxylic Acid-6-propylamides

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Abstract—SAR investigations of the 4- and 5-positions of a series of 4-amino-4*H*-pyran-2-carboxylic acid 6-carboxamides are reported. Potent inhibitors of influenza A sialidase with marked selectivity over the influenza B enzyme were obtained when the basic 4-amino substituent was replaced by hydroxyl or even deleted. Modifications at the 5-position exhibited a tight steric requirement, with trifluoroacetamide being optimal. © 2001 Elsevier Science Ltd. All rights reserved.

Introduction

Zanamivir 1¹ and GS 4071 2 (R=H)^{2,3} are potent selective inhibitors of influenza A and B sialidases and influenza A and B in vitro. The effectiveness of both zanamivir and the prodrug GS 4104 2 (R=Et) during clinical trials has demonstrated the utility of sialidase inhibitors as anti-influenza agents.⁴ Previously, as part of the SAR investigations into zanamivir, we reported on a series of 4*H*-pyran carboxamide influenza sialidase inhibitors 3.^{5,6} In these compounds the glycerol side chain of zanamivir was replaced by a lipophilic carboxamide moiety. Although some of this series were more potent inhibitors of influenza A sialidase than zanamivir, they were significantly less potent against the

influenza B enzyme. In this paper we describe further development of the SAR of the carboxamide series, looking at potency and selectivity. During these studies we have focused on the 4- and 5-positions on the pyran ring, structures 4 and 5, respectively. The in vitro and in vivo antiviral properties of representatives of these series are reported.

Results and Discussion

Although some of the previously described 4-amino-4*H*-pyran-2-carboxylic acid 6-carboxamides are potent inhibitors of influenza A sialidase, they are significantly

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less inhibitory of influenza B sialidase. 5,6 To expand the SAR studies of the carboxamide series, the 4- and 5-substituents were modified to investigate the effect on potency and selectivity.

Modification of the 4-position

Previous studies of the SAR of the 4-position of zanamivir suggested that modification of the 4-guanidino substituent was not tolerated. Also taking in consideration the potency of some of the 4-amino carboxamides¹⁴ so far described, we decided not to investigate 4-guanidino derivatives but to use amines 3a,b as our starting point. 4-Alkylamino derivatives 4a,b were synthesised using the route shown in Scheme 1. Whereas alkylation of the 4-amino derivative 6⁵ afforded gross mixtures, caesium carbonate mediated alkylation of the 4-trifluoroacetamide 7 gave good yields of the 4-N-alkylamino derivatives 8a,b. Deprotection under standard conditions⁵

yielded **4a,b**. ⁷ Using established chemistry ⁵ β-azide **9**⁸ was converted to the β-amino derivative **4c**.

The syntheses of the 4- α - and β -hydroxy epimers **4d** and **4e**, from the glycerol-protected 4- α - and β -hydroxy derivatives **10a**⁹ and **10b**, ¹⁰ respectively, are outlined in Scheme 2. The acid-stable *t*-butyldiphenylsilyl ether was used to protect the 4-hydroxy moiety of **10a** and **10b**, allowing the synthesis of acids **11a** and **11b**. TBTU¹¹ mediated coupling of acids **11a,b** with phenethyl-propylamine gave the corresponding amides in higher yields than the previously used two-step process via the pentafluorophenyl esters. ⁵

The 4-desoxy derivative **4f** was synthesised using a sodium borohydride reduction of the palladium complex of β -acetate **12** (Scheme 2).¹² Acetate **12** was obtained in 92% yield by acetylating the methyl ester of **4e** using acetic anhydride in pyridine.

$$(nC_3H_{7})_2N \xrightarrow{O} H$$

$$AcNH \xrightarrow{O} CO_2CHPh_2 \xrightarrow{a,b} AcNH \xrightarrow{O} CO_2CHPh_2 \xrightarrow{C} AcNH \xrightarrow{O} CO_2H$$

$$RHN \qquad 6: R = H \\ 7: R = CF_3CO \qquad 8b: R = CL_3 \\ AcO \xrightarrow{AcO} CO_2Me \qquad AcNH \xrightarrow{O} CO_2Me \qquad AcNH \xrightarrow{O} CO_2H$$

$$AcO \xrightarrow{AcO} CO_2Me \qquad AcNH \xrightarrow{O} CO_2Me \qquad AcNH \xrightarrow{O} CO_2H$$

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Scheme 1. (a) $(CF_3CO)_2O$ (93%); (b) Cs_2CO_3 , RBr, DMF (85% R=CH₃); (c) (i) TFA, CH₂Cl₂; (ii) K_2CO_3 , MeOH, H₂O (85% R=CH₃); (d) NaOMe, MeOH (94%); (e) (i) NaIO₄ MeOH, H₂O; (ii) NaClO₂, tBuOH (58%); (f) $CF_3CO_2C_6F_5$, pyridine; (g) $(nC_3H_7)_2NH$, THF (30% over 2 steps); (h) PPh₃, THF, then Et₃N, H₂O; (i) Et₃N, H₂O (74% over 2 steps).

Scheme 2. (a) $tBuPh_2SiCl$, DMAP, imidazole, DMF (77% 4-β-OH); (b) 80% aq AcOH (83% from 10a); (c) NaOMe, MeOH (99%); (d) (i) NaIO₄, MeOH, H₂O; (ii) NaClO₂, tBuOH (89% 4-β-OH); (e) TBTU, [Ph(CH₂)₂](nC_3H_7)NH, THF (42% 4-β-OH); (f) TBAF, THF (90% 4-β-OH); (g) Et₃N, H₂O (43% 4-β-OH); (h) Pd(Ph₃)₄, NaBH₄, THF (41%); (i) Et₃N, H₂O (69%).

The 4-N- α -methyl- and ethylamino derivatives **4a**,**b** (Table 1) exhibited at least a 10-fold loss of activity when compared to **3a**. These results mirrored the findings in the 6-glycerol series. ¹⁵ The 4- β -amino derivative **4c** proved to be approximately 10-fold less active than the corresponding 4- α -amino epimer **3a** both against influenza A and B sialidase.

To investigate whether the need for a basic group in the 4-position for potent activity still held for the carbox-amide series, we synthesised the epimeric $4-\alpha$ - and β -hydroxy derivatives **4d** and **4e**, respectively. Unexpectedly, the compounds exhibited similar activity against influenza A sialidase to **3b**, indicating this group was not as important for binding in this series as for

the 6-glycerol series. More surprisingly, the 4-desoxy derivative **4f** had similar activity to **3b** and **4d**. This was an important finding that could aid the design of further influenza sialidase inhibitors. Even against influenza B sialidase, where the carboxamide moiety is less favourably bound, removal of the basic amine of **3b** did not significantly affect the binding, as **4f** had a similar potency to **3b**. Overall, the modifications carried out did not reduce the difference in potency against influenza A and B and in some cases increased it.

Comparison of the crystal structures of **3b** and **4e** bound into influenza A sialidase (Fig. 1) or influenza B sialidase (data not shown) did not give any obvious rationale for the above observations, as the whole crystal

Table 1. Sialidase inhibitory and plaque reduction activities of 4a-f. Comparison with 1 and 3a,ba

	X	R	Sialidase inhibition IC ₅₀		Plaque reduction IC ₅₀	
			A Aichi (μM)	B Victoria (μM)	Flu A (μg/mL)	Flu B (μg/mL)
1			0.002	0.004	0.005	0.002
3a	α -NH ₂	<i>n</i> -Propyl	0.012	2.0	0.002	0.7
3b	α -NH ₂	$Ph(CH_2)_2$	0.003	3.6	0.03	0.28
4a	α- NHMe	n-Propyl	0.24	140		
4b	α-NHEt	n-Propyl	0.42	4.5		
4c	β -NH ₂	<i>n</i> -Propyl	0.14	7.4		
4d	α-ОН	$Ph(CH_2)_2$	0.007	51	0.005	>100
4e	β-ОН	$Ph(CH_2)_2$	0.017	>400		
4f	· Н	$Ph(CH_2)_2$	0.007	14		

^aSialidase inhibitory and plaque reduction activities of compounds were determined by previously reported methods.^{5,16}

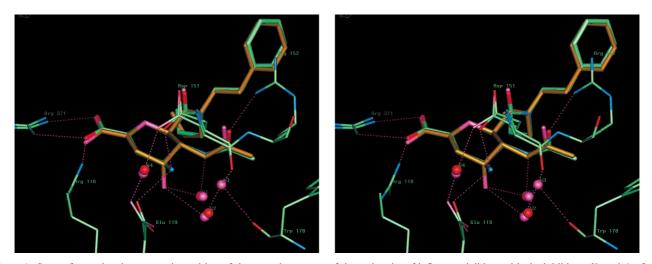


Figure 1. Stereo figure showing a superimposition of the crystal structure of the active site of influenza sialidase with the inhibitors 3b and 4e. Only the residues involved in interactions around the 4-position of the ligands are shown. Protein and ligand are in atom colours for 3b, ligand in yellow for 4e with oxygens of protein and water molecules in pink. Hydrogen bonds are shown as dotted lines.

5a-d: R = nPr, R₁ see Table 2 5e-h: R = $(CH_2)_2$ Ph, R₁ see Table 2

Scheme 3. (a) (i) NaIO₄, MeOH, H₂O; (ii) NaClO₂, tBuOH; (b) TBTU, Pr(R₁)NH, THF; (c) 4 M HCl in dioxan; (d) RCOCl or CH₃SO₂Cl, pyridine; (e) PPh₃, THF, then Et₃N, H₂O; (f) Et₃N, H₂O, 50 °C.

Table 2. Sialidase inhibitory and plaque reduction activities of 5a-h. Comparison with 3a,ba

	R	R_1	Sialidase inhibition IC ₅₀		Plaque reduction IC ₅₀	
			A Aichi (μM)	B Victoria (μM)	Flu A (μg/mL)	Flu B (μg/mL)
3a	n-Propyl	CH ₃ CO	0.012	2.0	0.002	0.7
5a	n-Propyl	C_2H_5CO	0.005	7.0	0.0002	3.0
5b	n-Propyl	(CH ₃) ₂ CHCO	17	>540		
5c	n-Propyl	CF ₃ CO	0.0003	0.2	0.00001	0.3
5d	n-Propyl	CH_3SO_2	3.0	92		
3b	$Ph(CH_2)_2$	CH ₃ CO	0.003	3.0	0.03	0.3
5e	$Ph(CH_2)_2$	C ₂ H ₅ CO	0.0007	6.0		
5f	$Ph(CH_2)_2$	C_3H_7CO	43	>500		
5g	$Ph(CH_2)_2$	cC ₃ H ₅ CO	0.05	90		
5h	$Ph(CH_2)_2$	CF ₃ CO	0.003	0.1	0.004	0.056

^aSialidase inhibitory and plaque reduction activities of compounds were determined by previously reported methods.^{5,16}

structures including the bound ligands were essentially superimposable. As can be seen in Figure 1 the only differences observed were slight movements of aspartic acid 151 and glutamic acid 119 and the accompanying structural water molecules near to the 4-position. These side-chain movements are due to the loss of hydrogen bonds to the amino group of **3b**. The small shifts of the water molecules occur as a consequence of the replacement of the α -amino group of **3b** with the β -hydroxy of **4e**.

The similarity of the activity of **3b** compared to **4c–f** suggests that within the carboxamide series the overall binding of the compounds is dominated by the lipophilic interactions of the carboxamide moiety. Therefore, any binding energy gained from hydrogen bonding of the 4-substituent with the enzyme and the structural water molecules is balanced by the desolvation of the native enzyme and unbound ligand.¹⁷

Modification of the 5-position

The synthesis of 5-modified derivatives **5a-f** from **13**¹³ is outlined in Scheme 3.

The activities of a range of 5-modified compounds are summarised in Table 2. Consistent with previous studies¹³ there was an extremely tight steric requirement for the 5-substituent. Replacement of acetamide with propionamide 5a,e was tolerated with little change in activity against influenza A or B sialidase. However, further homologation to either methylpropionamide 5b or 2-butyramide 5f resulted in a 1000-fold loss of activity. Remarkably, the cyclopropyl derivative 5g was only 25-fold less active than the acetamide 3b. The trifluoroacetamide derivative of the dipropylamide 5c was 100-fold more potent against influenza A in both enzyme and plaque assays. The 10-fold increase in potency in the influenza B enzyme assay did not translate into significantly improved influenza B plaque activity.

The trifluoroacetamide of the phenethyl-propylamide **5h** did not exhibit the large increase in influenza A activity shown by **5c**. However, the increase in the influenza B activity made **5h** the most potent 6-carboxamide derivative against influenza B. Replacement of the amide

moiety with a sulphonamide group **5d** gave a 300-fold loss of activity, whereas this modification was well tolerated in the 6-glycerol series.¹³

Conclusion

In contrast to the 6-glycerol substituted series related to zanamivir, the 6-carboxamide series does not require a basic group in the 4-position for potent sialidase activity. In fact, no functionality at all is required for nanomolar activity against influenza A sialidase. This surprising finding could be used to design further influenza sialidase inhibitors. Modifications at the 5-position demonstrated a very tight steric requirement for this substituent; trifluoroacetamide was optimal. As with the previous examples of the carboxamides, a significant selectivity for influenza A over influenza B sialidase was observed for all the compounds synthesised.

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References and Notes

- 1. Waghorn, S. L.; Goa, K. L. Drugs 1998, 55, 721.
- 2. Kim, C. U.; Lew, W.; Williams, M. A.; Zhang, L.; Liu, H.; Swaminathan, S.; Bischofberger, N.; Chen, M. S.; Tai, C. Y.; Mendel, D. B.; Laver, W. G.; Stevens, R. C. *J. Am. Chem. Soc.* **1997**, *119*, 681.
- 3. Kim, C. U.; Lew, W.; Williams, M. A.; Wu, H.; Zhang, L.; Chen, X.; Escarpe, P. A.; Mendel, D. B.; Laver, W. G.; Stevens, R. C. *J. Med. Chem.* **1998**, *41*, 2451.
- 4. Mendel, D. B.; Roberts, N. A. Curr. Opin. Infect. Dis. 1998, 11, 727.
- 5. Smith, P. W.; Sollis, S. L.; Howes, P. D.; Cherry, P. C.; Starkey, I. D.; Cobley, K. N.; Weston, H.; Scicinski, J.; Merritt, A.; Whittington, A.; Wyatt, P. G.; Taylor, N.; Green, D.; Bethell, R. C.; Madar, S.; Fenton, R. J.; Morley, P. J.; Pateman, T.; Beresford, A. J. Med. Chem. 1998, 41, 787.

- 6. Taylor, N. R.; Cleasby, A.; Singh, O.; Skarzynski, T.; Wonacott, A. J.; Smith, P. W.; Sollis, S. L.; Howes, P. D.; Cherry, P. C.; Bethell, R.; Colman, P.; Varghese, J. *J. Med. Chem.* **1998**, *41*, 798.
- 7. Satisfactory spectroscopic and analytical data were obtained for all new compounds.
- 8. Kok, G. B.; von Itzstein, M. Carbohydr. Res. 1997, 302, 237.
- 9. Okamoto, K.; Kaoru, T.; Goto, T. *Chem. Lett.* **1986**, 1449. 10. Schreiner, E.; Zbiral, E.; Kleineidam, R. G.; Schauer, R. *Liebigs Ann. Chem.* **1991**, 129.
- 11. Knorr, R.; Trzeciak, A.; Bannwarth, W.; Gillessen, D. Tetrahedron Lett. 1989, 30, 1927.
- 12. Keinan, E.; Roth, Z. J. Org. Chem. 1983, 48, 5302.
- 13. Smith, P. W.; Starkey, I. D.; Howes, P. D.; Sollis, S. L.;

- Keeling, S. P.; Cherry, P. C.; von Itzstein, M.; Wu, W. Y.; Jin, B. Eur. J. Med. Chem. 1996, 31, 143.
- 14. Chandler, M.; Bamford, M. J.; Conroy, R.; Lamont, B.; Patel, B. J. Chem. Soc., Perkin Trans. 1 1995, 1173.
- 15. von Itzstein, M.; Wu, W.-Y.; Phan, T. V.; Danylec, B.; Jin, B. PCT Int. Appl., WO 9116320, 1991.
- 16. Inhibition of influenza sialidase was determined in a fluorimetric assay by measuring the ability of compounds to inhibit the hydrolysis of 2'-(4-methylumbelliferyl)-α-D-N-acetylneuraminic acid (MUN) by whole virus (A/Aichi N2 or B Victoria) grown in hen eggs. The IC₅₀ value quoted is the concentration of inhibitor required to reduce the enzymic activity in this preparation by 50%.
- 17. Davis, A. M.; Teague, S. J. Angew. Chem., Int. Ed. 1999, 38, 736.